

involved in the transmission of mitogenic signals from tyrosine kinases to Ras (Pelicci G. et al Cell 70, 93-104 (1992)). The p52/46 Shc isoforms are involved in the cytoplasmic propagation of mitogenic signals from activated receptors to Ras (Bonfini L. et al Tibs 21, 257-261; 1996). They are rapidly phosphorylated on tyrosine after ligand stimulation of receptors and, upon phosphorylation, form stable complexes with activated receptors and Grb2, an adaptor protein for the Ras guanine nucleotide exchange factor SOS (Migliaccio E. et al Embo J. 16, 706-716 (1997); Pelicci G. et al Cell 70, 93-104 (1992); Rozakis-Adcock, M. et al Nature 360, 689-692 (1992)). These complexes induce Ras activation, as measured by increased RasGTP formation, Mitogen Activated Protein Kinase (MAPK) activity and FOS activation in cultured cells overexpressing p52^{shc}/p46^{shc} (Migliaccio, E. et al Embo J. 16, 706-716 (1997); Pronk, G. et al Mol.Cell. Biol. 14, 1575-1581 (1994); Lanfrancone, L. et al Oncogene 10, 907-917 (1995)). Likewise, p66^{shc} becomes tyrosine-phosphorylated upon receptor activation and forms stable complexes with activated receptors and Grb2. However, it inhibits c-fos promoter activation and does not affect MAPK activity, thereby suggesting that p66^{shc} acts in a distinct intracellular signalling pathway (Migliaccio E. et al Embo J. 16, 706-716 (1997)).

c-fos is transcriptionally activated in response to a large variety of adverse agents (environmental stress), such as DNA-damaging agents (e.g. ultraviolet radiation, UV) or agents that induce oxidative damage (e.g. hydrogen peroxide, H₂O₂) (Schreiber, M. et al Embo J. 14, 5338-5349 (1995); Sen, C. et al FASEB J. 10, 709-720 (1996)).

It is postulated that the major causal factor of aging is the accumulation of oxidative damage as an

organism ages (Martin, G. M. et al Nature Genetics 13, 25-34 (1996); Johnson, F.B. et al Cell 96, 291-302 (1999); Lithgow G. J. et al Science 273, 80 (1996)).

Indeed, transgenic flies that overexpress antioxidative enzymes have greater longevity (Orr W. C. et al Science 263, 1128-1130 (1994)); restriction of caloric intake lowers steady state levels of oxidative stress and damage and extends the maximum life span in mammals (Sohal R.S. et al Science 273, 59-63 (1996)). However, the genes that determine lifespan in mammals are not known. Among currently accepted evolutionary theories, it is postulated that aging is a post-reproductive process that has escaped the force of natural selection and that evolved through selection of alleles with early life benefits combined with pleiotropically harmful effects later in life. The postulated genes, since actively selected, are, therefore thought to regulate fundamental cellular processes, common to different species.

20 Summary of the Invention

The present inventors have determined that p66 is a pivotal gene in the regulation of the cellular responses to environmental and oncogenic stresses and that it is involved in the process of aging and in tumour suppression. p66 provides the first genetic information on the theory of aging. Mechanistically, p66 exerts its functions downstream to stress-activated serine kinases and upstream to p53-p21.

The present inventors have determined that targeted mutations of the mouse p66^{shc} gene induces stress resistance and prolongs survival. The present inventors disclose herein that i) p66^{shc} is serine phosphorylated upon UV treatment or oxidative damage; ii) the serine-

phosphorylation of p66 by oxidative signals is mediated by Erk1 and p38, as shown both *in vivo* and *in vitro*; iii) ablation of p66^{shc} expression by homologous recombination enhances resistance to oxidative damage both *in vitro* and *in vivo*; iv) a serine-phosphorylation defective mutant of p66^{shc} is unable to restore a normal stress response in p66^{shc} targeted cells; v) mice carrying the p66^{shc} targeted mutation have prolonged lifespan.

The present inventors disclose herein that i) p16, p53 and p21 activation is lost in p66^{-/-} cells upon H₂O₂ or UV treatment or RASV12 expression; ii) the oncogenic RASV12 is unable to induce cell senescence into p66^{-/-} mouse embryo fibroblasts (MEFs) and, on the contrary, it transforms p66^{-/-} cells; iii) p66^{-/-} MEFs over-expressing RASV12 show a transformed, spindle-shaped morphology, are capable of forming foci at confluency and colonies in semisolid media; and iv) p16 and p53 are unable to induce growth proliferation of p66^{-/-} cells.

Thus, the present inventors show herein that p66 itself is activated by serine phosphorylation by stress activated kinases and signals to p16-p19-p53-p21 and that functionally, the p66 signalling pathway regulates tumour suppression and lifespan.

Therefore, at its most general, the present invention provides materials and methods associated with the modulation of p66^{shc} gene expression and its involvement in a signal transduction pathway that is activated by environmental stresses and oncogenic

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mutations.

Thus, the present invention provides materials and methods for up-regulating or down regulating the amount of p66^{shc} polypeptide present in a cell. This regulation may be achieved at a nucleic acid level, or at a polypeptide level. These aspects are discussed below. However, primarily, the present invention provides a method of affecting the p66^{shc} signal transduction pathway in a cell, said method comprising the step of contacting said cell with an agent or substance capable of modulating p66^{shc} expression in said cell. The sort of agents preferred for such a method are discussed below.

In a first aspect of the present invention there is also provided a nucleic acid molecule comprising a p66^{shc} coding sequence incorporating at least one mutation as compared to the wild type sequence or the sequence as shown in Fig. 5 such that the protein encoded by the coding sequence has at least one serine residue absent or replaced by a different amino acid residue. Preferably, the serine residue is selected from the group consisting of S17, S19, S20, S26, S28, S36, S38, S40, S41, S54, S60, S66, S80, S120 and even more preferably selected from the group consisting of S28, S36 and S54. Even more preferably, the serine residue is replaced by a different amino acid residue, for example S36 is replaced by alanine (p66^{shc}S36A).

The nucleic acid of the present invention may comprises a p66^{shc} coding sequence which differs further from the wild type sequence or the sequence as shown in Fig. 5 in that it is a nucleic acid sequence that is an allele, mutant, variant or derivative, by way of nucleotide addition, insertion, substitution or deletion of the wild type sequence as illustrated in Fig. 5.

Systematic or random mutagenesis of nucleic acid to make an alteration to the nucleotide sequence may be performed using any technique known to those skilled in the art. In various embodiments of the present invention, a nucleic acid sequence that is a fragment, mutant, allele, derivative or variant, by way of addition, insertion or substitution or more or more nucleotides, of the p66^{shc} wild type sequence as illustrated in Fig. 5, has at least 60% homology, preferably at least 70% homology, more preferably at least 80% homology, more preferably at least 90% homology and even more preferably at least 95% homology.

In accordance with the present invention, nucleic acids having the appropriate level of sequence homology with the p66^{shc} nucleic acid sequence as shown in Fig. 5 may be identified by using hybridization and washing conditions of appropriate stringency. For example, hybridizations may be performed, according to the method of Sambrook et al., (22) using a hybridization solution comprising: 5X SSC, 5X Denhardt's reagent, 0.5-1.0% SDS, 100 µg/ml denatured, fragmented salmon sperm DNA, 0.05% sodium pyrophosphate and up to 50% formamide. Hybridization is carried out at 37-42°C for at least six hours. Following hybridization, filters are washed as follows: (1) 5 minutes at room temperature in 2X SSC and 1% SDS; (2) 15 minutes at room temperature in 2X SSC and 0.1% SDS; (3) 30 minutes-1 hour at 37°C in 1X SSC and 1% SDS; (4) 2 hours at 42-65°C in 1X SSC and 1% SDS, changing the solution every 30 minutes.

One common formula for calculating the stringency conditions required to achieve hybridization between nucleic acid molecules of a specified sequence homology is (Sambrook et al., 1989):

$T_m = 81.5^{\circ}\text{C} + 16.6\text{Log} [\text{Na}^+] + 0.41(\% \text{ G+C}) - 0.63 (\% \text{ formamide}) - 600/\text{\#bp in duplex}$

As an illustration of the above formula, using $[\text{Na}^+] = [0.368]$ and 50% formamide, with GC content of 42% and an average probe size of 200 bases, the T_m is 57°C . The T_m of a DNA duplex decreases by 1 - 1.5°C with every 1% decrease in homology. Thus, targets with greater than about 75% sequence identity would be observed using a hybridization temperature of 42°C . Such a sequence would be considered substantially homologous to the nucleic acid sequence of the present invention.

The present invention further provides a polypeptide encoded for by the nucleic acid molecule of the present invention as disclosed above. A preferred polypeptide comprises the p66^{shc} amino acid sequence or fragment thereof having at least one serine residue present in the wild type p66^{shc} sequence absent or replaced by a different amino acid residue. Preferably, the serine residue is one of S28, S36 or S54 and preferably, it is replaced with an alanine residue (e.g. p66^{shc}S36A).

Generally, nucleic acid according to the present invention is provided as an isolate, in isolated and/or purified form, or free or substantially free of material with which it is naturally associated, such as free or substantially free of nucleic acid flanking the gene in the human genome, except possibly one or more regulatory sequence(s) for expression. Nucleic acid may be wholly or partially synthetic and may include genomic DNA, cDNA or RNA. Where nucleic acid according to the invention includes RNA, reference to the sequence shown should be construed as reference to the RNA equivalent, with U substituted for T.

Nucleic acid sequences encoding all or part of the p66^{shc} gene and/or its regulatory elements can be readily

prepared by the skilled person using the information and references contained herein and techniques known in the art (for example, see Sambrook, Fritsch and Maniatis, "Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, 1989, and Ausubel et al, Short Protocols in Molecular Biology, John Wiley and Sons, 1992). These techniques include (i) the use of the polymerase chain reaction (PCR) to amplify samples of such nucleic acid, e.g. from genomic sources, (ii) chemical synthesis, or (iii) preparing cDNA sequences. Modifications to the p66^{shc} sequences can be made, e.g. using site directed mutagenesis, to lead to the expression of modified p66^{shc} polypeptide or to take account of codon preference in the host cells used to express the nucleic acid.

In order to obtain expression of the p66^{shc} nucleic acid sequences, the sequences can be incorporated in a vector having control sequences operably linked to the p66^{shc} nucleic acid to control its expression. The vectors may include other sequences such as promoters or enhancers to drive the expression of the inserted nucleic acid, nucleic acid sequences so that the p66^{shc} polypeptide is produced as a fusion and/or nucleic acid encoding secretion signals so that the polypeptide produced in the host cell is secreted from the cell. p66^{shc} polypeptide can then be obtained by transforming the vectors into host cells in which the vector is functional, culturing the host cells so that the p66^{shc} polypeptide is produced and recovering the p66^{shc} polypeptide from the host cells or the surrounding medium. Prokaryotic and eukaryotic cells are used for this purpose in the art, including strains of *E. coli*, yeast, and eukaryotic cells such as COS or CHO cells. The

choice of host cell can be used to control the properties of the p66^{shc} polypeptide expressed in those cells, e.g. controlling where the polypeptide is deposited in the host cells or affecting properties such as its glycosylation.

PCR techniques for the amplification of nucleic acid are described in US Patent No. 4,683,195. In general, such techniques require that sequence information from the ends of the target sequence is known to allow suitable forward and reverse oligonucleotide primers to be designed to be identical or similar to the polynucleotide sequence that is the target for the amplification. PCR comprises steps of denaturation of template nucleic acid (if double-stranded), annealing of primer to target, and polymerisation. The nucleic acid probed or used as template in the amplification reaction may be genomic DNA, cDNA or RNA. PCR can be used to amplify specific sequences from genomic DNA, specific RNA sequences and cDNA transcribed from mRNA, bacteriophage or plasmid sequences. The p66^{shc} nucleic acid sequences provided herein readily allow the skilled person to design PCR primers. References for the general use of PCR techniques include Mullis et al, Cold Spring Harbor Symp. Quant. Biol., 51:263, (1987), Ehrlich (ed), PCR technology, Stockton Press, NY, 1989, Ehrlich et al, Science, 252:1643-1650, (1991), "PCR protocols; A Guide to Methods and Applications", Eds. Innis et al, Academic Press, New York, (1990).

An oligonucleotide for use in nucleic acid amplification may have about 10 or fewer codons (e.g. 6, 7 or 8), i.e. be about 30 or fewer nucleotides in length (e.g. 18, 21 or 24). Generally specific primers are upwards of 14 nucleotides in length, but not more than

18-20. Those skilled in the art are well versed in the design of primers for use processes such as PCR.

5 A further aspect of the present invention provides an oligonucleotide or polynucleotide fragment of the nucleotide sequence shown in Fig. 5 or a complementary sequence, in particular, for use in a method of obtaining and/or screening nucleic acid. The sequences referred to above may be modified by addition, substitution, insertion or deletion of one or more nucleotides, but
10 preferably without abolition of ability to hybridize selectively with nucleic acid with the sequence as shown in Fig. 5.

In some preferred embodiments, oligonucleotides according to the present invention that are fragments of
15 the sequence shown in Fig. 5, are at least 10 nucleotides in length, more preferably at least 15 nucleotides in length, more preferably about 20 nucleotides in length. In most cases, oligonucleotides according to the present invention are no greater than 2 kb, preferably no greater
20 than 1.5 kb, more preferably no greater than 1 kb and even more preferably no greater than 0.5 kb. Such fragments themselves individually represent aspects of the present invention. Fragments and other oligonucleotides may be used as primers or probes as
25 discussed but may also be generated (e.g. by PCR) in methods concerned with determining the presence in a test sample of a p66^{hsc} gene sequence.

A convenient way of producing a polypeptide according to the present invention is to express nucleic acid encoding it, by use of the nucleic acid in an
30 expression system. The use of expression system has reached an advanced degree of sophistication today.

Accordingly, the present invention also encompasses

a method of making a polypeptide (as disclosed), the method including expression from nucleic acid encoding the polypeptide (generally nucleic acid according to the invention). This may conveniently be achieved by growing a host cell in culture, containing such a vector, under appropriate conditions which cause or allow expression of the polypeptide. Polypeptides may also be expressed in vitro systems, such as reticulocyte lysate.

Systems for cloning and expression of a polypeptide in a variety of different host cells are well known. Suitable host cells include bacteria, eukaryotic cells such as mammalian and yeast, and baculovirus systems. Mammalian cell lines available in the art for expression of a heterologous polypeptide include Chinese hamster ovary cells, HeLa cells, baby hamster kidney cells, COS cells and many others. A common, preferred bacterial host is *E. coli*.

Suitable vectors can be chosen or constructed, containing appropriate regulatory sequences, including promoter sequences, terminator fragments, polyadenylation sequences, enhancer sequences, marker genes and other sequences as appropriate. Vectors may be plasmids, viral e.g. 'phage, or phagemid, as appropriate. For further details see, for example, *Molecular Cloning: a Laboratory Manual*: 2nd edition, Sambrook et al., 1989, Cold Spring Harbor Laboratory Press. Many known techniques and protocols for manipulation of nucleic acid, for example in preparation of nucleic acid constructs, mutagenesis, sequencing, introduction of DNA into cells and gene expression, and analysis of proteins, are described in detail in *Current Protocols in Molecular Biology*, Ausubel et al. eds., John Wiley & Sons, 1992.

Thus, the present invention further provides a host

cell containing nucleic acid as disclosed herein. The nucleic acid of the invention may be integrated into the genome (e.g. chromosome) of the host cell. Integration may be promoted by inclusion of sequences which promote recombination with the genome, in accordance with standard techniques. The nucleic acid may be on an extra-chromosomal vector within the cell.

Both the nucleic acid molecules and the polypeptides as disclosed herein may be used in a method of treatment and in particular may be used in the preparation of a medicament for increasing the cellular resistance to oxidative stress.

Therefore, the present invention also provides methods of increasing resistance in cells to oxidative stress. Such oxidative stress may be as a result of external, e.g. environmental, factors such as UV, X-rays heat shock, osmotic shock, oxidative stress (singlet oxygen, H_2O_2 , hydroxylradicals, inflammatory cytokines). or it may be as a result of internal factors resulting in necrosis of cells as occurs in some disease states.

A method of increasing resistance to oxidative stress may comprise disrupting a $p66^{shc}$ signalling pathway. The pathway may be disrupted at any stage during the signalling process, for example, the $p66^{shc}$ polypeptide may be mutated such that the serine residue is absent or replaced by a different amino acid residue, e.g. alanine such that the resulting polypeptide cannot be serine phosphorylated; the ability of molecules such as p38 or MAPK to phosphorylate $p66^{shc}$ may be disrupted by, for example, dominant negative kinases or specific inhibitors; and, most preferably, the expression of $p66^{shc}$ may be disrupted thereby down regulating $p66^{shc}$ polypeptide in the cell. Further, as p53 and p16 are not

biologically active in p66^{shc} cells, any dominant negative p66 molecules may be used to block p16 and p53 function.

The disruption of p66^{shc} gene expression may be obtained in various ways. Antisense oligonucleotide sequences based on the p66^{shc} sequence may be designed to hybridise to the complementary sequence of nucleic acid, pre-mRNA, or mature mRNA, interfering with the production of polypeptide encoded by a given DNA sequence (e.g. either native p66^{shc} polypeptide or a mutant form thereof), so that its expression is reduced or prevented altogether. In addition to the p66^{shc} coding sequence, antisense techniques can be used to target the control sequences of the p66^{shc} gene, e.g. in the 5' flanking sequence of the p66^{shc} coding sequence, whereby the antisense oligonucleotides can interfere with the p66^{shc} control sequences. The construction of antisense sequences and their use is described in Peyman and Ulman, Chemical Reviews, 90:543-584, (1990), Crooke, Ann. Rev. Pharmacol. Toxicol., 32:329-376, (1992), and Zamecnik and Stephenson, P.N.A.S., 75:280-284, (1974).

In a second aspect, the present invention provides a method of screening for compounds capable of modulating a p66^{shc} signalling pathway comprising contacting a candidate compound with a p66^{shc} expression system as described above; determining the amount of a component of the signalling pathway; and comparing said amount of the component with the amount of the component in the absence of said candidate compound.

Preferably, the expression system comprises a nucleic acid vector having a p66^{shc} coding sequence or fragment thereof inserted therein. Suitable vectors can be chosen or constructed, containing appropriate

regulatory sequences, including promoter sequences, termination fragments, polyadenylation sequences enhancer sequences, marker genes and other sequences as appropriate. For further details see, for example, 5 Molecular cloning: A laboratory Manual: 2nd edition, Sambrook et al, 1989, Cold Spring Harbor Laboratory Press. Procedures for introducing nucleic acid into cells depends on the host cell used, but are well known.

Thus the expression system may also comprises a host 10 cell containing a p66^{shc} coding sequence or a vector as disclosed above. Most preferably, the expression system comprises a cell derived from a cell line, such as mouse embryo fibroblasts, known to express p66^{shc}.

The p66^{shc} signalling pathway may be modulated, e.g. 15 disrupted, at any stage during the signalling pathway such that production of active p66^{shc} is prevented. Examples of such modulation may include directly preventing expression of p66^{shc} by blocking factors involved in the transcription of genes. For example, 20 antisense primers may be used to bind to the p66^{shc} gene thereby preventing transcription factors binding to regulatory agents required for promoting transcription. Alternatively, the coding sequence for p66^{shc} may be targeted so as to introduce mutations which prevent the 25 expression of p66^{shc} without disrupting expression of associated proteins such as P52^{shc} and p46^{shc}. Preferably, the mutation disrupts the exon encoding the p66 CH2 region. Antisense probes may also be used for binding DNA or mRNA encoding p66^{shc} such that its translation is 30 prevented. Alternatively, antibodies specific for p66^{shc} (e.g. anti-CH2 antibodies) may be used to specifically bind to the expressed protein such that subsequent binding of p66^{shc} to other proteins, e.g. receptors, is

prevented.

Further, inhibition of the p66 function can be obtained by inhibiting the phosphorylation of the p66 CH2 region induced by Erk 1 or p38 stress-kinases. To this end, library of compounds can be screened to identify those which are able to inhibit the in vitro phosphorylation of the GST-CH2 region by recombinant Erk1. Examples of such compounds include MAPK (mitogen activated protein kinase), any serine/threonine kinase, and p38.

Thus, the present invention provides use of p66^{shc} nucleic acid or polypeptides for screening for molecules which affect or modulate its activity or function. Such molecules may be useful in a therapeutic (possibly including prophylactic) context and include nucleic acid sequence (antisense or sense sequences capable of specifically binding to p66^{shc} nucleic acid sequences (RNA or DNA, antibodies (monoclonal or polyclonal), and protein nucleic acids (PNA).

Antibodies may also be provided by the present invention for affecting or modulating the activity or function of p66^{shc}. Thus, a further and important use of the p66^{shc} polypeptide is in the raising of antibodies that have the property of specifically binding to the p66^{shc} polypeptide, fragments or active portions thereof.

The production of monoclonal antibodies is well established in the art. Monoclonal antibodies can be subjected to the techniques of recombinant DNA technology to produce other antibodies or chimeric molecules which retain the specificity of the original antibody. Such techniques may involve introducing DNA encoding the immunoglobulin variable region, or the complementarity determining regions (CDRs), of an antibody to the

constant regions, or constant regions plus framework regions, of a different immunoglobulin. See, for instance, EP-A-184187, GB-A-2188638 or EP-A-239400. A hybridoma producing a monoclonal antibody may be subject to genetic mutation or other changes, which may or may not alter the binding specificity of antibodies produced.

Accordingly, a further aspect of the present invention provides an antibody able to bind specifically to the p66^{Shc} polypeptide. Such an antibody may be specific in the sense of being able to distinguish between the polypeptide it is able to bind and other human polypeptides for which it has no or substantially no binding affinity (e.g. a binding affinity of about 1000x worse). Specific antibodies bind an epitope on the molecule which is either not present or is not accessible on other molecules. Antibodies according to the present invention are preferably specific for the wild-type p66^{Shc} polypeptide such that they disrupt its function. Alternatively, these specific antibodies may simply be used as a tool for identifying the p66^{Shc} polypeptide in a biological sample. In this regard, the antibody may be labelled in order to aid detection.

Preferred antibodies according to the invention are isolated, in the sense of being free from contaminants such as antibodies able to bind other polypeptides and/or free of serum components. Monoclonal antibodies are preferred for some purposes, though polyclonal antibodies are within the scope of the present invention.

Antibodies may be obtained using techniques which are standard in the art. Methods of producing antibodies include immunising a mammal (e.g. mouse, rat, rabbit, horse, goat, sheep or monkey) with the protein or a fragment thereof. Antibodies may be obtained from

immunised animals using any of a variety of techniques known in the art, and screened, preferably using binding of antibody to antigen of interest. For instance, Western blotting techniques or immunoprecipitation may be used (Armitage et al, Nature, 357:80-82, 1992). Isolation of antibodies and/or antibody-producing cells from an animal may be accompanied by a step of sacrificing the animal.

As an alternative or supplement to immunising a mammal with a peptide, an antibody specific for a protein may be obtained from a recombinantly produced library of expressed immunoglobulin variable domains, e.g. using lambda bacteriophage or filamentous bacteriophage which display functional immunoglobulin binding domains on their surfaces; for instance see WO92/01047. The library may be naive, that is constructed from sequences obtained from an organism which has not been immunised with any of the proteins (or fragments), or may be one constructed using sequences obtained from an organism which has been exposed to the antigen of interest.

Antibodies according to the present invention may be modified in a number of ways. Indeed the term "antibody" should be construed as covering any binding substance having a binding domain with the required specificity. Thus the invention covers antibody fragments, derivatives, functional equivalents and homologues of antibodies, including synthetic molecules and molecules whose shape mimics that of an antibody enabling it to bind an antigen or epitope.

Example antibody fragments, capable of binding an antigen or other binding partner are the Fab fragment consisting of the VL, VH, Cl and CH1 domains; the Fd fragment consisting of the VH and CH1 domains; the Fv

fragment consisting of the VL and VH domains of a single arm of an antibody; the dAb fragment which consists of a VH domain; isolated CDR regions and F(ab')₂ fragments, a bivalent fragment including two Fab fragments linked by a disulphide bridge at the hinge region. Single chain Fv fragments are also included.

It is well known that pharmaceutical research leading to the identification of a new drug may involve screening of very large numbers of candidate substances, both before and even after a lead compound has been found. This is one factor which makes pharmaceutical research very expensive and time consuming. Means for assisting in the screening process can have considerable commercial importance and utility. Such means for screening for substances potentially useful in increasing resistance to oxidative stress and thus extending cellular longevity, is provided by the present invention.

A method for screening for a substance which modulates (disrupts) activity of the p66^{shc} polypeptide may include contacting one or more test substances with the polypeptide in a suitable reaction medium, testing the activity of the treated polypeptide and comparing that activity with the activity of the polypeptide in comparable reaction medium untreated with the test substance or substances. Conveniently, an enzyme activity assay may be used to determine any disruption in the activity of p66^{shc}.

Combinatorial library technology provides an efficient way of testing a potentially large number of different substances for ability to disrupt or delete activity of p66^{shc}. Such libraries and their use are known in the art. The use of peptide libraries is preferred.

Further, assays for determining p66 inhibitors may

include usage of wildtype cells (both established cell lines or primary cells capable of expressing p66^{shc}, e.g. MEFs, p66-/- MEFs or MEFs overexpressing p66 (through the usage of a p66^{shc} expressing vector).

- 5 Comparative responses to be determined may include response to stress factors, e.g. UV or H₂O₂; inhibition of RASV12 (or any other oncogene) -induced senescence in primary fibroblasts; inhibition of p53 or p19 or p16 or p21 function (as measured by transcriptional assays, or
- 10 stability assays, or nucleus-cytoplasmic export assays); or inhibition of p66 phosphorylation induced by RASV12 or by oxidative stress signals.

- Following identification of a substance or compound which disrupts p66^{shc} or a step in the p66^{shc} signalling
- 15 pathway, the substance or compound may be investigated further. Furthermore, it may be manufactured and/or used in the preparation, i.e manufacture or formulation, of a composition such as a medicament, pharmaceutical composition or drug. These may be administered to
- 20 individuals. Oxidative stress has been implicated in the generation of many human diseases. Thus, medicaments, pharmaceutical compositions or drugs comprising an inhibitor of p66 function, or substances or compounds which disrupt p66^{shc} or a step in the p66^{shc} pathway may be
- 25 used in the treatment of diseases such as arteriosclerosis; ischemic heart disease, Parkinson, Alzheimer, vascular complications of diabetes, emphysema and other lung diseases, myocardial infarction, stroke, premature aging, cell senescence, skin diseases and
- 30 cancers.

The present invention also provides a method of increasing cellular resistance to oxidative stress comprising deleting or disrupting the gene encoding p66^{shc}

from the cellular genome. Such a method may include processes such as gene therapy wherein a nucleic acid vector comprises a nucleic acid sequence capable of being incorporated into the genome of the cell and disrupting the expression of p66^{shc}.

Vectors such as viral vectors have been used in the prior art to introduce genes into a wide variety of different target cells. Typically the vectors are exposed to the target cells so that transfection can take place in a sufficient proportion of the cells to provide a useful therapeutic or prophylactic effect from the expression of the desired polypeptide. The transfected nucleic acid may be permanently incorporated into the genome of each of the targeted tumour cells, providing long lasting effect, or alternatively the treatment may have to be repeated periodically.

A variety of vectors, both viral vectors and plasmid vectors, are known in the art, see US Patent No. 5,252,479 and WO 93/07282. In particular, a number of viruses have been used as gene transfer vectors, including papovaviruses, such as SV40, vaccinia virus, herpes viruses, including HSV and EBV, and retroviruses. Many gene therapy protocols in the prior art have used disabled murine retroviruses.

As an alternative to the use of viral vectors other known methods of introducing nucleic acid into cells includes electroporation, calcium phosphate co-precipitation, mechanical techniques such as microinjection, transfer mediated by liposomes and direct DNA uptake and receptor-mediated DNA transfer.

As mentioned above, the aim of gene therapy using nucleic acid encoding the p66^{shc} polypeptide, or an active portion thereof, is to increase the amount of the

expression product of the nucleic acid in cells in which the level of the wild-type p66^{abc} polypeptide is absent or present only at reduced levels or where it would be advantageous to express an modified p66^{abc} polypeptide having improved properties over those of the wild type. Such treatment may be therapeutic in the treatment of cells which are already effected by oxidative stress or prophylactic in the treatment of individuals susceptible to oxidative stress and aging.

The present invention further provides genetic markers for aging. Such markers may provided materials and methods for determining a predisposition to aging associated with certain disease states. As disclosed herein, heterozygous p66+/- mice have a slight, but evident, increase in longevity, thereby suggesting that slight variations in the expression levels of p66 may influence aging. This may provide for the determination of variations in the p66 RNA transcriptional regulatory sequences (e.g. the promoter) which affect the p66 transcription rate and longevity. Also, since the lifespan correlates with the functional activity of p66, there may be allelic variations in the p66 coding sequence that also correlate with different length of lifespan.

As mentioned above, the present inventors have also determined that p66 expression is indispensable for the capacity of p53 to induce apoptosis (following, for example, oxidative stress) or senescence (following for example, Ras expression). Thus, agonists of p66 can be envisioned as putative tumour suppressors. In fact, any molecule that would prevent p66 de-phosphorylation or kinases that induce p66 phosphorylation are potential devices for tumour treatment.

Thus, in a further aspect of the present invention, there is provided methods of increasing resistance to tumour formation by increasing expression of P66^{shc}. The results disclosed herein indicate that increased levels of P66^{shc} reduce susceptibility to carcinogenesis. Therefore, the present invention further includes the use of P66^{shc} (nucleic acid molecules or polypeptides as disclosed herein) for reducing susceptibility to cancers. Likewise, the present invention provides use of P66^{shc} in the preparation of a medicament for the treatment and prevention of cancers.

P66^{shc} for such use, may be in the form of a polypeptide or may be in the form of a nucleic acid molecule which encodes a functional P66^{shc} polypeptide. The nucleic acid may be in the form of an expression vector which comprises a nucleic acid molecule encoding part or all of P66^{shc} polypeptide. The expression vector may be used as part of a gene therapy application as disclosed herein. Further, in this aspect, agents may be used which increase the expression of P66^{shc} within the cell. Such agents may be further nucleic acid molecules or compounds such as transcription factors, which are capable of increasing the expression of P66^{shc} or compounds that increase the levels of p66 expression acting at post-transcriptional levels, such as at the level of stability of RNA or protein.

For example, nucleic acid encoding authentic biologically active p66^{shc} polypeptide could be used in a method of gene therapy, to treat a patient who is unable to synthesize the active polypeptide or unable to synthesize it at the normal level, thereby providing the effect provided by wild-type p66^{shc} and suppressing the susceptibility to tumour formation. Of course, gene

therapy may be used to boost the levels of p66^{shc} expression or alternatively, nucleic acid encoding a modified p66^{shc} polypeptide may be used in order to improve the effects of p66^{shc} expression.

5 Thus, the present invention further provides a method of screening for substances that modulates (increases/improves) activity of the p66^{shc} polypeptide. Such a method may include contacting one or more test substances with the polypeptide in a suitable reaction medium, testing the activity of the treated polypeptide and comparing that activity with the activity of the polypeptide in comparable reaction medium untreated with the test substance or substances. Alternatively, increased/improved p66^{shc} activity may be achieved by 10 increasing p66^{shc} gene expression in vitro or in vivo. An expression system as described above may be used to screen for substances or compounds (agonists) that are capable of increasing expression of p66^{shc} nucleic acid.

Aspects and embodiments of the invention will now be 20 illustrated, by way of example, with reference to the accompanying figures. Further aspects of the invention will be apparent to those skilled in the art. All documents mentioned in this text are incorporated herein by reference.

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Brief Description of the Drawings

In the figures

Figure 1 shows serine phosphorylation of p66^{shc} by UV or H₂O₂ treatment. Figure 1A: Modular organisation of 30 p66^{shc}; Y: Y239, Y340 and Y317, the major Shc tyrosine phosphorylation sites. The alternative initiation codon (ATG) of p46^{shc} is indicated. Figure 1B: Anti-phosphotyrosine Western blotting of anti-p66 (α CH2)

immunoprecipitates from lysates of serum starved MEFs (SF) or Mouse Embryo Fibroblasts (MEFs) treated with EGF, UV or H_2O_2 for 5 min or 4 hrs, as indicated. The same blot was reprobed with anti-p66^{shc} antibodies (α -CH2). The p66^{shc} polypeptides are arrowed. Immunoglobulin cross-reactive polypeptides are also indicated (Ig). Figure 1C: Western blotting analysis of p66^{shc} expression of serum starved MEFs (SF) or MEFs treated with EGF, UV or H_2O_2 for 5 min or 4 hrs as indicated. The same blot was reprobed with anti-actin antibodies. Figure 1D: Phosphoaminoacid analysis of p66^{shc}. Serum-starved MEFs (SF) were labelled with 1mCi/ml [³²P] orthophosphate for 4 hr and unstimulated (SF) or cells stimulated with EGF, UV or H_2O_2 for 5 min or 4 hr were lysed and immunoprecipitates were resolved by SDS-PAGE and transferred to nitrocellulose and autoradiographed (not shown). Phosphoaminoacid analysis was performed on the p66^{shc} polypeptide. Positions of the phosphoserine (S), phosphothreonine (T) and phosphotryosine (Y) markers are indicated.

Figure 2 shows p66^{shc} enhances stress oxidative response *in vitro* and *in vivo*. Figure 2A: Western blotting analysis of Shc expression in p66^{shc}+/+ or p66^{shc}-/- MEFs (left panel) and in p66^{shc}-/- MEFs transduced with vector alone, p66^{shc} or p66^{shc}S36A cDNAs (right panel). Figure 2B: Viability of MEFs after H_2O_2 treatment. Equal numbers (1.2×10^5) of the indicated MEF cells grown in 100mm dishes in triplicate were infected with the PINCO retrovirus or with PINCO retroviruses expressing p66^{shc} or p66^{shc}S36A, as indicated, kept for additional 48 hrs to allow gene expression of exogenous cDNAs and exposed to 400mM H_2O_2 for 24 hrs. Cell viability was determined by trypan blue exclusion. Results are expressed as a percentage of viable cells with respect to H_2O_2 untreated

controls and represent the mean of three independent experiments. Expression of p66^{shc} or p66^{shc}S36A did not significantly influence viability or growth rate of MEFs, as measured 48 hrs after viral infection (not shown).

5 **Figure 3** shows mapping of p66^{shc} serine-phosphorylation sites. Figure 3A: Anti-CH2 western blots of lysates from MEFs transfected with vectors expressing the isolated CH2 region and then starved (SF) or treated with EGF or UV for 5 min or 4 hr, as indicated. The star
10 indicates the shifted CH2 polypeptide. Figure 3B: The S36A or S54A mutations were introduced with the isolated CH2 region or the full-length p66^{shc}. The resulting cDNAs were HA-tagged, cloned with a pcDNA3 expression vector and transfected into MEFs. Cultures were treated as
15 indicated and analysed by western blotting using anti-HA antibodies. Figure 3C: Phosphoaminoacid analysis of p66^{shc} and p66^{shc}S36A. MEFs were transfected with HA-p66^{shc} or HA p66^{shc}S36A expression vectors, kept in culture for 48 hrs labelled with 1mCi/ml [³²P] orthophosphate for 4 hr and
20 treated with EGF or H₂O₂ for 5 min, as indicated, lysed and immunoprecipitated with anti-HA antibodies. Phosphoaminoacid analysis was performed on the HA-p66^{shc} or HA-p66^{shc}S36A polypeptides, as described in Fig. 1 legend.

25 **Figure 4** shows cumulative survival (Kaplan and Meier) or p66^{shc}+/+ (dashed line), p66^{shc}/+/- (dotted line) and p66^{shc}-/- (solid line) mice. Survival of the p66^{shc}-/- mice was 71.4%.

30 **Figure 5** shows (a) the p66 cDNA nucleotide sequence (the ATG initiation site is underlined) and (b), separated, the p66 amino acid sequence.

Figure 6 shows a 13kb genomic region containing all the Shc coding exons which was characterised by

restriction enzyme mapping and nucleotide sequence of all exon-intron boundaries and 5' regulatory regions.

Figure 7 shows the construction of the targeting vector pBSp66ShcKO.

Figure 8 shows the vector pBSp66ShcKO with a TK transcriptional unit cloned at its 3' end.

Figure 9 shows multiple metabolic changes in p66^{Shc}-/- cells. In particular it shows the mechanism of p66 action and its metabolic effects, in particular the increased production of H₂O₂ and the consequent increase in concentration of Reactive Oxygen Species (ROS); and the increased production of ATP (at the cost of ROS). p66 is also confirmed to be located in the mitochondrial membrane and regulates a transition port and is involved in electron transfer.

Figure 10 shows regulation of mitochondrial activity by p66^{Shc}.

Figure 11 shows the role of p66 in the p53-dependent apoptosis.

Detailed Description

1) Cell lines, reagents and plasmid construction

Mouse embryo fibroblasts (MEFs) were isolated from 12 to 14 day embryos derived from p66^{Shc}-/- mice and p66^{Shc}+/+ mice and maintained in Earle's minimal essential medium supplemented with 10% fetal bovine serum. The S36A and S54A mutations were generated by standard PCR techniques. The p66^{Shc}, p66^{Shc}S36A, HA-CH2, HA-CH2S35A, HA-CH2S54A, HA-p66^{Shc}, HA-p66^{Shc}-S35A and HA-p66^{Shc}S54A were cloned into the pCDNA3 or PINCO eukaryotic expression vectors (Claudio P.P. et al Cancer Res. 54, 5556-5560 (1994); Grignani, F. et al Cancer Res. 1, 14-19 (1998)).

The antibodies used were: the anti-Shc polyclonal antibody which recognises the SH2 domain of all three Shc isoforms (Pelicci G. et al Cell 70, 93-104 (1992)); the anti p66 polyclonal antibody which recognises the p66^{shc} isoform (Migliaccio E. et al Embo J. 16, 706-716 (1997)); the anti- β -actin polyclonal antibody, (Sigma Immuno Chemicals); the anti-HA monoclonal antibody; the anti-phosphotyrosine monoclonal antibody, (Santa Cruz Biotechnology).

2) Metabolic labeling immunoprecipitation, Western blotting and phosphoaminoacid analysis.

For whole lysates, cells were directly lysed in SDS sample buffer (50mM Tris-HCl pH 6.8, 2% SDS v/v, 10% glycerol and 5% v/v β -mercaptoethanol) and boiled for 5 min. 50 μ g of total protein was analysed by SDS-PAGE. For immunoprecipitation, cells were lysed on ice in PY buffer (20mM Tris-HCl pH 7.8, 50mM NaCl, 30mM Na₂P₂O₇, 5mM sodium orthovanadate, 1% v/v Triton x-100 containing freshly added protease inhibitors: 1mM phenylmethyl sulfonyl fluoride, 10 μ gml⁻¹ leupeptin and 5 mg ml⁻¹ aprotinin), appropriate antibodies were adsorbed on Protein A Sepharose (Pharmacia) and then incubated with cell lysates for 2hr at 4°C. Immunoprecipitates were recovered, resolved by 10% SDS-PAGE and transferred to nitrocellulose filters, as described elsewhere (Migliaccio E. et al Embo J. 16, 706-716 (1997)). Blots were blocked, probed with specific antibodies and immune complexes revealed by horseradish-peroxidase conjugated with specific secondary antiserum (Biorad) followed by enhanced chemiluminescence. For phosphoaminoacid analysis, cells were grown to confluence on 10 cm plates, starved in serum-free medium and labelled for 4h in 5 ml

phosphate free DMEM containing 5% dialyzed FBS and 1mCi ml^{-1} ^{32}P -orthophosphate. Cells were stimulated with 30ng ml^{-1} EGF or $400\mu\text{M H}_2\text{O}_2$, or irradiated with 50 J/m^2 UV, rinsed twice with ice cold PBS and lysed in PY buffer.

5 p66^{shc} proteins were isolated by immunoprecipitation with anti-SHC or anti-HA antibodies and resolved by SDS-PAGE. p66^{shc} polypeptides were transferred to PVDF membranes and hydrolyzed in 6M HCl for 60 min at 110°C . The hydrolysis products were separated in the presence of phosphoserine, 10 phosphothreonine and phosphotyrosine markers by SDS-PAGE at pH1.9 and pH3.5 in two dimensions on TLC plates.

3) Transfections, infections and cellular viability test.

MEFs were transfected with the LipofectAMINE PLUS 15 Reagent GibcoBRL (average transfection efficiency 50%). For retroviral infections, the empty PINCO vector and recombinant PINCO vectors expressing p66^{shc} or p66^{shc}S36A cDNAs were transfected into the phoenix amphotropic packaging cell line and, after 48 hrs, supernatants were 20 used to infect MEFs cells (1.2×10^5 cells/100mm dish). The efficiency of infection (GFP positive cells) was determined by FACS analysis 48 hr after infection. Viability was assessed by the trypan-blue dye exclusion test.

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4) Statistical analysis

Survival functions were estimated by the Kaplan and Meier product limit method. Survival distributions were compared using the logrank test (Marubini E. et al New 30 York, John Wiley & Sons (1995). All statistical calculations were performed using SAS/STAT Rel. 6.12 software (SAS Institute 1995).

5) Construction of p66^{Shc} targeting vector and electroporation and selection of ES cells.

The targeting vector was constructed using standard cloning procedures. The plasmid was linearized with Kpn 1 before electroporation into ES cells. CJ7 ES cells were maintained on a monolayer of mitomycin C inactivated, neomycin-resistant primary embryonic fibroblast. A suspension of 15 million trypsinized ES cells in PBS was electroporated with 25 μ g of DNA of the linearized targeting vector by using the Bio-Rad gene pulser II apparatus with 240 V and 500 μ F. Cells were plated immediately after transfection and allowed to recover for 24 hr before selection in medium 350 μ g/ml Geneticin and 2 μ M of ganciclovir. Cells were fed daily and after 9 days the resulting colonies were picked and cultivated singularly until extraction and freezing.

6) Southern blot analysis of ES cells and Mice.

To identify the mutated Shc allele, genomic DNA from ES cells and from the mouse tails was prepared by proteinase K lysis and phenol-chloroform extraction, digested with Eco R1 and analyzed by Southern blot analysis. A 1.1.Kb Eco R1-Xba 1 fragment was used as external probe to discriminate between the WT 8 kb and the recombinant 3.5 kb allele bands (Fig 2).

7) Generation of mice carrying the disrupted p66 Shc allele.

Two different clones of targeted ES were used to generate chimeric mice. C57BL/6J blastocysts injected with 10-15 ES cells were transferred to pseudopregnant female mice. Chimeric mice, identifiable by agouti coat color, were mated with C57BL/6J mice. Offspring with

agouti coat color were tested for the presence of the recombined allele by means of Southern blot analysis. Heterozygotes obtained from the crosses of the chimeras with 129Sv female mice were interbred to establish the colony of p66 $-/-$ mice in the 129 genetic background. The mice were housed at a constant room temperature (22°C) and humidity (60%) with a 12 h light/dark cycle, with free access to standard mouse chow and tap water.

10 Results

1) Construction of the p66shc targeting vector.

The present inventors have mutated the mouse Shc locus using conventional embryonic stem cell technology. The genomic Shc locus was isolated from a 129 mouse genomic library. A 13 Kb genomic region containing all the Shc coding exons was characterized by restriction enzyme mapping and nucleotide sequence of all exon-intron boundaries and 5' regulatory regions (Fig. 6). A positive/negative (G418/Ganciclovir) selection strategy was applied to introduce a mutation in the region of first coding exon that contains the p66 ATG. To construct the targeting vector (pBSp66ShcK0; Fig 7) we used the EcoR1 8 KB fragment containing exons 1-8 (Fig.7). The Bal 1 fragment containing the p66 specific start site (Fig.7) was substituted with the Neo transcriptional unit driven by the pY promoter. The resulting vectors contains 2,2 and 4,6 Kb flanking sequences at the 5' and 3' ends, respectively (Fig. 7). A TK transcriptional unit was cloned at its 3' end (Fig.3).

2) ES tranfection and selection.

CJ7 ES cells, provided by Dr. Vera Soares (Memorial Sloan Kettering Cancer Center, NY, USA) were transfected by electroporation, and resistant colonies were selected and screened for the deletion of the p66 first coding sequence by Southern blotting. The screening strategy was based on the EcoRI site introduced in the targeted allele by the insertion of the Neo transcriptional unit (Fig.7) 24 ES clones of 150 analyzed showed one targeted Shc allele. Cytogenetic analysis of 9 ES clones revealed a normal modal number of chromosomes.

3) Generation of targeted mice.

Two ES clones were injected into C57BL/6J blastocysts, according to established procedures. Breeding of heterozygous ($p66^{sch+/-}$) yielded the expected frequency of homozygous animals. Analysis of the genotype of the animals was performed by Southern blotting of DNA extracted from the tails and confirmed by Western blotting of $p66^{shc}$ expression in various tissues.

4) p66 is phosphorylated on Ser36 after H_2O_2 or UV stimulation.

Since Fos is transcriptionally activated by a variety of environmental stresses (hydrogen peroxide: H_2O_2 ; ultraviolet irradiation: UV), the present inventors have analysed the modifications of p66 upon H_2O_2 or UV stimulation of mouse and human fibroblasts. Results revealed that p66 is markedly phosphorylated on serine upon H_2O_2 /UV irradiation. Further, the present inventors mapped the major serine phosphorylation site to Ser 36. A Ser-Ala 36 p66 mutant is not phosphorylated by UV/ H_2O_2 in vivo.

5) **Erk1 and p38 mediates UV/H₂O₂ phosphorylation of p66**

By using purified enzymes in in vitro kinase assays and either dominant negative kinases or specific inhibitors in vivo, the present inventors have demonstrated that p66 is phosphorylated by p38 and Erk1 upon UV or H₂O₂ stimulation.

Erk 1 (also known as MAPK), JNK and p38 are stress-induced kinases. To identify the kinase(s) responsible for the phosphorylation of p66 induced by oxidative stress in vivo, the present inventors analysed the extent of p66 phosphorylation after stress signals in MEFs expressing a JNK dominant negative kinase or in cells treated with various MAPK and p38 specific inhibitors [PD98059, which prevents activation of Erk 1 by Raf; SB203580 which specifically inhibits the p38 Map kinases]. Results showed that SB203580 and PD98059, but not the JNK dominant negative kinase, prevented p66 phosphorylation by oxidative stress signals (H₂O₂), indicating that p66 is phosphorylated in vivo by Erk1 and p38, but not by JNK.

The present inventors then reconstructed the p66 phosphorylation in vitro by using recombinant Erk1 or JNK and the bacterially expressed p66 CH2 region (CH2 was expressed in bacteria as GST-fusion protein). Erk1, but not JNK, was unable to in vitro phosphorylate the p66 CH2 region. Phosphorylation was specific, as shown by the finding that, in the same assay, Erk1 was unable to phosphorylate the p66 CH2 region when the S36A mutation was introduced.

6) **p66 modulates the oxidative stress response in vitro**

H₂O₂ treatment induces fibroblast cell death. The present inventors have demonstrated that: i) overexpression of p66 in wild-type MEFs increases cell

death induced by H_2O_2 ; ii) p66^{-/-} MEFs are more resistant to H_2O_2 -induced cell death than wild-type controls *in vivo*. Paraquat is a pesticide that kills mice by inducing oxidative damage. The present inventors have further demonstrated that p66^{-/-} mice are more resistant to paraquat treatment than littermates.

7) p66 regulates the p16, p53 and p21 response

Since environmental stresses activates the p16 - p53-p21 signalling pathways, the present inventors have further investigated whether p66 interferes with p16-p53-p21 activation by H_2O_2 . Results revealed that p16, p53 and p21 activation are lost in p66^{-/-} cell upon H_2O_2 treatment.

8) p66 is a tumour supressor

In vitro, the stimulatory effect of p66 on the p53-p21 pathway suggests that it might play a role in the cellular response on oncogenic stimuli. Therefore, the present inventors have evaluated the effects of p66 on the response of primary fibroblasts on the oncogenic RASV12 mutant. RASV12 induces senescence of wild-type MEFs, as a consequence of p53-p21 activation. Expression of RASV12 into p66^{-/-} MEFs induced cellular transformation. The present inventors have demonstrated that p53 and p16 are unable to induce senescence of mouse p66^{-/-} fibroblasts.

9) p66 mediates aging

The results presented herein demonstrate that p66 is involved in the cellular response to stresses

(environmental and oncogenic). The present inventors therefore considered whether p66 is also involved in mediating aging. To do this, they evaluated the survival of p66^{-/-} mice. Of the many mice that were born in August 1996, 14^{+/+}, 8^{+/-} and 15^{-/-} were not sacrificed and kept for survival analysis. Evaluation after 28 months (December 1998) revealed

+/+	0/14 survivors	0%
+/-	3/8 survivors	37%
-/-	11/15 survivors	73%

10) P66^{-/-} MEFs are resistant to p53-induced apoptosis

To analyse the molecular mechanisms underlying the increased resistance of p66^{-/-} MEFs to oxidative stress-induced apoptosis, the present inventors analysed the function of p53 in p66^{-/-} MEFs (versus wt MEFs) following H₂O₂ treatment.

P53 is a tumour suppressor gene that functions as a transcription factor. P53 is also a critical component of cellular mechanisms that respond to a variety of different environmental stresses, including DNA damage, hypoxia or oxidative stress, and which lead to cell cycle arrest or apoptosis. Stress-induced p53 activation involves increased protein stability and post-translational modifications including phosphorylation and acetylation.

H₂O₂ treatment induced up-regulation of p53 into wt as well as p66^{-/-} MEFs, implying that the mechanisms that underlie increased p53 stability upon oxidative stress are not affected by p66 expression. However, overexpression of p53 (using retroviruses or adenoviruses) into p66^{-/-} MEFs was unable to induce

apoptosis, indicating that the p53-downstream pathway is altered in the absence of p66.

To understand the mechanisms underlying the loss of p53 activity into p66-/- cells, the inventors measured the potential of p53 to act as a transcription factor in p66-/- cells. To this end, they performed transactivation experiments using p53 and a p53-target promoter into p66-/- and wt MEFs. Results showed that the transcriptional activity of p53 is retained into p66-/- MEFs. This finding suggest that the inability of p53 to execute the apoptotic process into p66-/- MEFs might be due to alterations of the activity of (some) p53-target genes.

Little is known on the p53-target genes which are involved in the execution of the many p53 biological activities (apoptosis, senescence, cell cycle arrest). In the case of apoptosis, it has been recently demonstrated that p53 activate a set of genes (so called PIGs; p53 inducible genes) that are involved in the control of ROS metabolism. Indeed, p53 activation results in the transcriptional activation of these genes, increased intracellular concentrations of ROS, caspase activation and apoptosis.

To test whether this intracellular pathway is altered into p66-/- MEFs, the inventors have measured the intracellular concentration of ROS into p66-/- and wt MEFs following p53 overexpression (using fluorescent dyes, such as the DCFDA). Results showed that the p53-induced peak of intracellular ROS following p53 expression is lost into the p66-/- MEFs.

Together, these results indicate that p66 regulates ROS metabolism and is a crucial effector of p53-mediated apoptosis.

11) P66-/- MEFs are resistant to Ras-induced replicative senescence

p53 regulates a checkpoint response to oncogenic signals: expression of oncogenic mutants of Ras in primary cells activate p53, resulting in premature senescence, whilst the absence of functional p53 is sufficient for oncogenic transformation of murine fibroblasts. P53, indeed, is mutated at high frequency in many tumour types, implying that its action is central to tumour development. The mechanisms through which cells sense hyperproliferation and which lead to p53 activation and cellular senescence remain, however, poorly understood. It has been recently demonstrated that Ras-induced senescence involved generation of ROS, implying that ROS are also involved in the execution of the senescence program by p53.

Therefore, The inventors have analysed whether Ras is able to induce cellular senescence into p66-/- MEFs. Results showed that p66-/- MEFs are resistant to Ras-induced senescence, suggesting that the intracellular mechanisms that regulate the response to oncogenic signals is altered into p66-/- cells.

To investigate whether p66-/- MEFs are more susceptible to tumour formation, the present inventors analysed the ability of Ras-expressing p66-/- MEFs to form colonies into methylcellulose (a property of transformed cells), as compared to p53-/- MEFs. While Ras expression induced transformation of p53-/- MEFs, it was unable to do so into p66-/- MEFs. Likewise, p66-/- mice did not show increased frequency of spontaneous or UV (or TPA)-induced tumours.

The inventors conclude that p66 expression is indispensable for the capacity of p53 to induce apoptosis

(following oxidative stress) or senescence (following Ras expression). However, the loss of p66 does not mimic the loss of p53 with respect to the tumour-prone phenotype, suggesting that other pathways are activated by p53 following oncogenic signals.

Potential applications.

p66 is a downstream target of p53 in the execution of the p53-mediated apoptosis and cellular senescence. Manipulations of p66, therefore, can be used to restore the p53 function into p53 null cells. A potential application of this finding is the possibility to activate the apoptotic or senescence pathways in tumour with non-functional p53 (the p19-p53 pathways are non-functional in the vast majority of tumours). Agonist of p66 can be envisioned as putative tumour suppressors. Any molecule that would prevent p66 de-phosphorylation or kinases that induce p66 phosphorylation are potential device for tumour treatment.

12) The intracellular concentration of ROS (reactive oxygen species) in p66^{-/-} MEFs is decreased.

The above-mentioned results suggest that p66 is involved in the regulation of ROS metabolism. Therefore the inventors have measured the intracellular concentrations of ROS in cultures of wt and p66^{-/-} MEFs (using fluorescent dyes, such as the DCFDA). Results showed that the intracellular concentration of ROS is consistently decreased in p66^{-/-} MEFs (of about one third).

Potential implications.

Oxidative stress has been implicated in the

generation of many human diseases. ROS induce damage to a variety of macromolecules, such as proteins (disulphide bond formation); lipids (peroxidation); DNA (mutations). These alterations are thought to mediate the phenotypic traits of aging and to be implicated in the pathogenesis of different diseases (atherosclerosis; ischemic heart disease; Parkinson, Alzheimer, vascular complications of diabetes). Inhibitors of the p66 function might be used in the treatment of these diseases.

13) P66 is a mitochondrial protein .

The present inventors have investigated the mechanisms underlying the regulation of ROS production by p66. ROS are mainly produced by mitochondria, as a consequence of the minimal, yet physiological, uncoupling of the electron transfer during oxidative phosphorylation. Therefore, the inventors have first evaluated whether p66 is a mitochondrial protein. Elector microscopic, immunofluorescence and cell fractionation studies (using anti-p66 specific polyclonal and monoclonal antibodies) revealed that p66 is a mitochondrial protein, located in the inner membrane of the mitochondria.

14) The decrease of the mitochondrial $\Delta\psi$ ($m\Delta\psi$) following H2O2 treatment is slowed in p66-/- MEFs .

To assess if the different survival response to H2O2 treatment of p66-/- MEFs also reflects a different mitochondrial response, the inventors tested the functionality of the mitochondria in the p66-/- and WT MEFs after treatment with H₂O₂.

The mitochondrial electric potential ($m\Delta\psi$) is a direct measure of the electro-chemical gradient developed

by the activity of the electron transfer chain complexes and the resulting proton pumping. $m\Delta\psi$ was, therefore, measured as indicator of mitochondrial activity after H_2O_2 treatment, using tetramethylrhodamine methyl ester (TMRM). TMRM is a lipophilic cationic fluorescent probe which is accumulated in the mitochondria according to the Nerst's equation. The higher is the $m\Delta\psi$, the more intensive is the signal resulting from the fluorescence of the TMRM, thereby allowing determination of $m\Delta\psi$ variations in cell culture systems.

The inventors found that the reduction of the $m\Delta\psi$ after 10 minutes from the addition of 400 μM of H_2O_2 is about 50% in the wt MEFs and only 20% in the p66-/- MEFs. This result indicates that mitochondria in p66-/- are less damaged by H_2O_2 with respect to those of wt cells.

15) Isolated mitochondria from p66-/- mouse livers are more coupled following stimulation with different substrates

In order to determine if mitochondrial respiratory performances are altered by the absence of the p66 protein, purified mitochondria from WT and p66-/- mouse livers were analysed for their respiratory activity. In vitro quantitation of oxygen consumption by the mitochondrial suspensions was performed using an oxygen sensitive electrode. The inventors measured both basal and stimulated respiration following administration of ADP, succinate, aspartate, malate, dinitrophenol, and calculated the state 3 / state 4 ratio of WT and p66-/- mitochondria. The inventors consistently found higher values of this ratio for the p66-/- as compared to the wt mitochondria. These findings suggest that mitochondria extracted from p66-/- livers are in a better "coupled

state".

16) Swelling as a measure of Permeability Transition induced by H_2O_2 is lowered in isolated p66-/- respect to the WT mouse liver mitochondria .

Mammalian mitochondria possess an inner membrane channel, the permeability transition pore (MTP). Opening of the MTP is responsible for the permeability transition (PT), a sudden increase in permeability to solutes with molecular masses approximately 1500 Da (experimentally obtained after Ca^{2+} accumulation in the presence of a variety of so-called "inducing agents"). The PT is the primary event in apoptosis, and, together with mitochondrial depolarization, it represents the "central executioner" in the apoptotic cascade. PT results in a modification of the mitochondrial volume and in a consequent swelling of the organelle.

It is possible to analyse the relative MTP opening probability and the resulting PT by measuring the variation of absorbance of the swollen using a spectrophotometer at 540 nm wavelength. The inventors analysed the "swelling" response of isolated liver mitochondria from WT and p66-/- mice after different PT stimuli. An impairment/delay of the swelling was detected in the p66-/- mitochondria after treatment with $100 \mu M H_2O_2$. This finding indicates that the MTP in the p66-/- mitochondria is less sensitive to oxidation and its consequent opening induced by H_2O_2 suggesting that the regulation of the opening of the MTP is one of the events controlled by p66 in the mitochondria. p66, therefore, would function favouring the opening of the pore, thereby determining a more fragile and uncoupled mitochondria. This would result into less efficient respiratory

performance, more active production of ROS and better apoptotic response to oxidative stress.

Potential implications

5 Since p66 controls the function of the permeability pore in mitochondria and, indirectly, the efficiency of electron transfer and ROS production, alterations of the p66 function are expected to allow manipulation of metabolism and apoptosis.

17) Old Lung cellular density is higher in the p66-/-

10 All levels of organisation of an organism, from molecules to complete organ systems, undergo senescence and a large body of evidence indicates that free radicals are the primary cause of this phenomenon. The resulting oxidative stress induced aging is a process strictly associated with the decrease of functional capacities in all parts of the body and even if it does not clearly define the causes of death occurring at the end of the life span of an organism, it is generally charged to this degenerative senile syndrome.

20 Some studies suggest that kidney and heart failure are the most common cause of death at very old age in humans as in other mammals but also disfunctions in other organs due to aging significantly contribute to the death at the end.

25 In particular, in the lung, the volume/weight ration of inflated dried lung parenchyma increase significantly with aging resulting in the so-called "aging-lung emphysema" or "senile emphysema". The oxidative stress accumulated during aging induced programmed cell death which may contribute to the thinning of the alveolar septa observed with aging. The examination of aged (more

then 1 year old) p66-/- and wild type mouse lungs reveals that p66-/- lung weights and sizes are larger than wild type. Histological analysis of these lungs shows a general pattern of higher cellular density and in particular thicker alveolar walls are present in the p66-/- lung.

The present inventors, therefore have concluded that the delayed thinning of the alveolar walls and the consequent early appearance of signs of senile-emphysema in wild type mice could suggest a possible mechanistic explanation which may be extended to other organs, for the prolonged life observed in the absence of p66.

Potential implications

Regardless of the contribution of delayed senile emphysema to the increased lifespan of p66-/- mice, these findings indicate that the lung is a target organ for p66 functions. Inhibition of p66 function into the lung, may therefore lead to increased functionality of the lung and provide new treatments for emphysema.

Discussion

To investigate its role in the cellular stress response, the present inventors analysed the extent of p66^{shc} tyrosine-phosphorylation in mouse embryo fibroblasts (MEFs) treated with UV or H₂O₂, as compared to the effects of treatment with growth factors, such as the epidermal-growth factor (EGF). Anti-p66 immunoprecipitates from lysates of untreated and EGF-, UV-, or H₂O₂-stimulated fibroblasts were immunoblotted with anti-phosphotyrosine antibodies. EGF stimulation induced a marked increase in the phosphotyrosine content of p66^{shc}, which was maximal after 5 min. Neither UV nor

H₂O₂ treatment induced significant tyrosinephosphorylation of p66^{shc} (Fig.1B). However, Western blotting of the same lysates with anti-Shc antibodies revealed a marked gel retardation of the p66^{shc} polypeptides after 5 min and 4 hours treatment with UV or H₂O₂, consistent with other post-translational modifications of p66^{shc} induced by these agents (Fig.1C). Therefore, p66^{shc} phosphorylation was analyzed by phosphoaminoacid analysis (Fig.2C). p66^{shc} polypeptides from serum-starved cells were phosphorylated primarily on serine. UV (Fig.2C) and H₂O₂ (not shown) induced a marked increase in the level of phosphoserine and had no effect on phosphotyrosine. In contrast, EGF induced a marked increase in the level of phosphotyrosine and a modest increase in phosphoserine (Fig.2C). It appears, therefore, that p66^{shc} is involved in the intracellular transduction pathways of both environmental stresses and growth factors, albeit with distinct functions, since UV and H₂O₂ induced rapid and persistent serine-phosphorylation, while EGF induced rapid and transient tyrosine-phosphorylation.

To investigate the functional role of p66^{shc} in the stress oxidative response, the present inventors next analysed the effects of p66^{shc} overexpression or p66^{shc} ablation on the cellular response of MEFs to H₂O₂. MEFs were derived from mice carrying a targeted mutation of the Shc locus that disrupted the exon encoding the p66 CH2 region, without affecting the p52^{shc}/p46^{shc} coding sequences (M. Gioglio et al : submitted for publication). p66^{shc}/+ MEFs from wild-type mice with otherwise identical genetic background were used for comparison. As expected, expression of p66^{shc} was normal in p66^{shc}/+ MEFs while was undetectable in p66^{shc}-/- MEFs; expression p52^{shc}/p46^{shc} was identical in p66^{shc}/+ and p66^{shc}-/- MEFs.

p66^{shc}+/+ MEFs were susceptible to H₂O₂ treatment, with more than 70% of cells being killed after 24 hours exposure to 400 μ M H₂O₂. Overexpression of p66^{shc} rendered p66^{shc}+/+ more susceptible to H₂O₂ treatment (approximately 85-90% cell death after 24 hours). In contrast, p66^{shc}-/- MEF cells were more resistant to killing by the same dose of H₂O₂ and more than 70% of these cells survived after 24 hours of H₂O₂ treatment (Fig.2B). Expression of the p66^{shc} cDNA into p66^{shc}-/- cells restored a normal response to H₂O₂ (Fig.2B).

The present inventors then examined the ability of p66^{shc}-/- mice to resist oxidative stress *in vivo*. To this end, mice were treated with paraquat, which, upon intake by the cell, generates superoxide anion. At a dosage of 70mg/kg, 5 of 5 p66^{shc}+/+ mice died within 48 hours after paraquat administration. In contrast, out of 5 p66^{shc}-/- treated mice, two died within the first 48 hours, two after approximately 72 hours and 1 survived for several weeks (Fig.2C). Together, these results point to a function of p66^{shc} in the cellular stress oxidative response.

To investigate whether p66^{shc} participates in the cellular stress response as a cytoplasmic transducer of stress signals, The present inventors analysed the potential of a serine-phosphorylation defective mutant of p66^{shc} to rescue the impaired stress oxidative response of p66^{shc}-/- MEFs. The p66^{shc} CH2 region probably contains the p66^{shc} major serine-phosphorylation site(s), as suggested by the gel-mobility shift induced by H₂O₂ and EGF, when the CH2 was expressed in cultured cells as isolated domain (Fig.3A). The CH2 region contains three serine residues with a consensus sequence for serine/threonine kinase phosphorylation (S28, S36 and S54) (Davis, J. J.

Biol. Chem. 268, 14553-14556 (1993)). Alanine substitution of S36 abrogated the gel-mobility shift of both isolated CH2 and full-length p66^{shc} induced by H₂O₂ (Fig.3B). Phosphoaminoacid analysis of the p66^{shc} and p66^{shc}S36A polypeptides revealed a marked increase in the level of phosphoserine induced by H₂O₂ in the p66^{shc}, but not in the p66^{shc}S36A mutant (Fig.3C), thereby confirming that S36 is the p66^{shc} major serine phosphorylation site.

The present inventors then expressed the p66^{shc}S36A mutant into p66^{shc}-/- MEFs and evaluated its effect on the stress oxidative response. As shown in Fig. 2B, p66^{shc}S36A was unable to restore a normal response to H₂O₂. Instead, it conferred further resistance to H₂O₂-induced cell death, probably through a dominant negative effect on the stress response-signalling pathway. Together, these results indicate that p66^{shc} acts as a signal transducer in the cellular response to oxidative stress.

Enhanced resistance to environmental stress correlates with prolonged lifespan in invertebrates. In *S.Cerevisiae*, deletions of RAS1 (Sun, J. et al J. Biol. Chem. 269, 18638-18645 (1994); Kale, S. P. et al Dev. Genet. 18, 154-160 (1996)) or mutations in the SIR4 (Kennedy, B. K. et al Cell 80, 485-496 (1995)) locus increase lifespan and resistance to starvation, ethanol and heat shock or UV, respectively. In *C. elegans*, mutants of genes of the dauer signalling pathway, such as age-1 and daf-2, survive longer and are more resistant to oxygen radicals, heat and UV (Murakami, S. et al Genetics, 143, 1207-1218 (1985); Larsen, P. L. et al Genetics 139, 1576-1583 (1995)). In *D.melanogaster*, selection for late-life fitness is associated with greater resistance to environmental stresses (Service, P. M. et al Physiol, Zool. 58, 380-389 (1985); Service P.M.

Physiol Zool. 60, 321-326 (1987); Arking R. et al Dev. Genet. 12 362-370 (1991)), and hypomorphic mutants of the *mth* locus live 35% longer and are more resistant to dietary paraquat and starvation (Lin, Y. J. et al Science 282, (1998). The present inventors, therefore, retrospectively analysed the effects of the $p66^{shc}$ mutation on lifespan. 37 mice born on August 1996 from $p66^{shc+/-}$ heterozygous parents were not sacrificed and maintained under identical conditions of stability. They consisted of 14 $p66^{shc+/-}$, 8 $p66^{shc+/-}$ and 15 $p66^{shc-/-}$ mice. After 28 months of observation, all the wild-type animals had died (median survival of 25.37 ± 0.63 months), while 3 of the 8 heterozygous (37%) and 11 of the 15 homozygous (73%) were still alive. The remaining 3 $p66^{shc+/-}$ died after additional two months (median survival of 27.40 ± 2.819). 3 $p66^{shc-/-}$ mice also died after two months; the remaining 9 are still alive (lifespan more than 31 months). The comparison of survival curves obtained by the Kaplan and Meier method (Marubini, E. et al Valsecchi, M.G. New York, John Wiley & Sons (1995)) (Fig.4) showed a highly significant difference between the three groups (log-rank $p=0.0002$). Cumulative survival did not differ significantly between wild type and heterozygous ($p=ns$ [0.057]). The cumulative survival in the $p66^{shc+/-}$ group was 71.4% ($p<0.01$ vs $p66^{shc+/-}$ and $p66^{shc-/-}$). Therefore, it appears that homozygous mutation of $p66^{shc}$ correlates with prolonged survival in mice.

The results presented herein are consistent with a model in which lifespan is determined as a result of the increased ability to resist or repair environmental damage. $p66^{shc}$ is part of a signal transduction pathway, which is activated by environmental stresses (H_2O_2 or UV) and whose mutation increases stress resistance and

lifespan. Biochemical and genetic investigation of the p66^{shc} signalling pathway should lead to better understanding of mechanisms relevant to aging in mammals.

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Claims:

1. A nucleic acid molecule comprising a P66^{shc} coding sequence incorporating at least one mutation as compared to the wild type sequence or the sequence as shown in Fig. 5 such that the protein encoded by the coding sequence has at least one serine residue absent or replaced by a different amino acid residue.
2. A nucleic acid molecule according to claim 1 wherein the serine residue is selected from the group S17, S19, S20, S26, S28, S36, S38, S40, S41, S54, S60, S66, S80 or S120.
3. A nucleic acid molecule according to claim 1 or claim 2 wherein the serine residue is selected from the group S28, S36 and S54.
4. A nucleic acid molecule according to any one of the preceding claims wherein the serine residue is S36 and is replaced by alanine (p66^{shc}S36A)
5. A polypeptide encoded by a nucleic acid molecule according to any one of the preceding claims.
6. A replicable vector comprising nucleic acid according to any one of claims 1 to 4 operably linked to control sequences to directs its expression.
7. A host cell transformed with a vector according to claim 6.
8. A method of producing a modified p66^{shc} polypeptide comprising culturing a host cell according to claim 7 so

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that the p66^{shc} polypeptide is produced.

9. A method of affecting the p66^{shc} signal transduction pathway in a cell, said method comprising the step of
5 contacting said cell with an agent capable of modulating p66^{shc} gene expression.

10. A method according to claim 9 wherein said agent is a nucleic acid molecule capable of hybridizing to nucleic acid encoding p66^{shc} thereby reducing or preventing said
10 p66^{shc} thereby reducing or preventing said p66^{shc} expression.

11. A method according to claim 9 wherein said agent is a vector comprising nucleic acid encoding p66^{shc}, said
15 vector being capable of incorporating said nucleic acid into the genome of the cell so that the nucleic acid encoding p66^{shc} is expressed in the cell.

12. A method of increasing resistance in cells to oxidative stress comprising the step of disrupting the
20 p66^{shc} signalling pathway.

13. A method according to claim 12 wherein said step of
25 disrupting the p66^{shc} effects the susceptibility of p66^{shc} to phosphorylation.

14. A method according to claim 12 or claim 13 wherein said step of disrupting the p66^{shc} pathway causes a mutant
30 p66^{shc} polypeptide to be expressed such that at least one serine residue present in the wild type p66^{shc} is absent or replaced by a different amino acid residue.

15. A method according to claim 14 wherein said serine

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residue is S36 and is replaced by alanine

16. A method according to claim 14 wherein said mutant polypeptide cannot be serine phosphorylated.

17. A method according to any one of claims 12 to 16 wherein said disruption effects the ability of a serine/threonine kinase, p38 or MAPK to phosphorylate p66^{shc}.

18. A method according to claim 12 wherein the step of disrupting the p66^{shc} signalling pathway includes contacting the cell with an antibody binding domain capable of specifically binding to the p66^{shc} polypeptide such that its function is disrupted or prevented.

19. A method according to claim 12 wherein said step of disrupting the p66^{shc} signalling pathway includes disrupting the p66^{shc} gene expression.

20. A method according to claim 19 wherein disruption of the p66^{shc} gene expression includes contacting the cell with a substance capable of interfering with the expression of nucleic acid encoding the p66^{shc} polypeptide so as to reduce or prevent its production.

21. A method according to claim 20 wherein the substance is an antisense oligonucleotide capable of hybridising to the nucleic acid encoding the p66^{shc} polypeptide.

22. Use of a substance which disrupts p66^{shc} or a step in the p66^{shc} signalling pathway, in the preparation of a medicament to increase cellular resistance to oxidative stress.

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23. Use of an antisense oligonucleotide capable of specifically hybridising to p66^{shc} nucleic acid in the preparation of a medicament for increasing resistance in cells to oxidative stress.

5

24. Use according to claim 23 wherein said antisense oligonucleotide is RNA

25. Use according to claim 23 or claim 24 wherein the p66^{shc} nucleic acid sequence is shown in Fig. 5.

10

26. Use of an antibody binding domain capable of specifically binding to a p66^{shc} polypeptide or fragment thereof in the preparation of a medicament for increasing resistance in cells to oxidative stress.

15

27. Use according to any one of claims 22 to 26 wherein the medicament is for the treatment of diseases including lung emphysema, myocardial infarction, stroke, premature aging, cell senescence, Parkinson's, Alzheimer, cancers and diabetes.

20

28. A method of increasing resistance to tumour formation in a tissue comprising the step of increasing the expression of p66^{shc} in said tissue.

25

29. A method according to claim 28 wherein the step of increasing the expression of p66^{shc} includes contacting the tissue with an agent capable of increasing expression of p66^{shc} gene.

30

30. A method according to claim 29 wherein said agent is a transcription factor.

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31. A method according to claim 29 wherein said agent is a vector comprising nucleic acid encoding p66^{shc} polypeptide said vector being capable incorporating said nucleic acid into the genome the cells of the tissue.

5

32. A method of screening for compounds capable of modulating a p66^{shc} signalling pathway comprising contacting a candidate compound with a p66^{shc} expression system; determining the amount of a compound of the signalling pathway; and comparing said amount of the component with the amount of the component in the absence of said candidate compound.

10

15

33. A method according to claim 32 further comprising the step of preparing a pharmaceutical composition comprising the candidate compound capable of modulating a p66^{shc} pathway and a pharmaceutical acceptable carrier.

20

34. A method according to claim 32 or claim 33 wherein said step of determining the amount of a compound of the signalling pathway is an enzyme activity assay.

25

35. A method according to any one of claims 32 to 34 wherein said candidate compounds include nucleic acid sequences, antibody binding domains, and protein nucleic acids.

30

36. A method of reducing intracellular levels of reactive oxygen species (ROS) in a cell, said method comprising the step of contacting said cell with an agent capable of inhibiting the expression or activity of p66^{shc} polypeptide.

37. A method according to claim 36 wherein said agent is

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a nucleic acid molecule capable of specifically hybridising with nucleic acid with the cell which codes for the p66^{shc} polypeptide such that expression the p66^{shc} polypeptide is reduced or prevented.

5

38. A method according to claim 36 wherein the agent is an antibody binding domain capable of specifically binding to the p66^{shc} polypeptide such that its functions are inhibited or prevented.

10

39. Use of an oligonucleotide sequence capable of specifically hybridising to a p66^{shc} nucleic acid coding sequence or fragment thereof for detecting the presence or absence of p66^{shc} nucleic acid in a biological sample.

15

40. Use according to claim 39 wherein said oligonucleotide is more than 20 nucleotides in length and is derived from the sequence shown in Fig. 5,

20

41. Use of an antibody binding domain capable of specifically binding to a p66^{shc} polypeptide for detecting the presence or absence of p66^{shc} polypeptide in a biological sample.

25

42. A method of determining the presence or absence of a p66^{shc} nucleic acid or a mutant, variant derivative or allele thereof in a biological sample, comprising the step of contacting said sample with a nucleic acid molecule capable of hybridising specifically with said p66^{shc} nucleic acid or a mutant, variant derivative or allele thereof and determining whether or not hybridization has taken place.

30

43. A method of determining the presence or absence of a

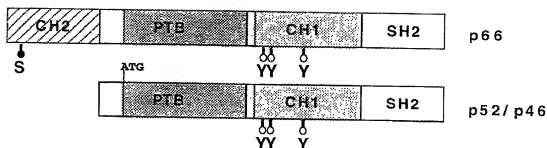
- 54 -

- p66^{shc} polypeptide or a mutant, variant derivative or allele thereof in a biological sample, comprising the step of contacting said sample with an antibody binding domain capable of hybridising specifically with said
- 5 p66^{shc} nucleic acid or a mutant, variant derivative or allele thereof and determining whether or not hybridization has taken place.
44. An expression system comprising a nucleic acid
- 10 vector having a p66^{shc} coding sequence or fragment thereof inserted therein.

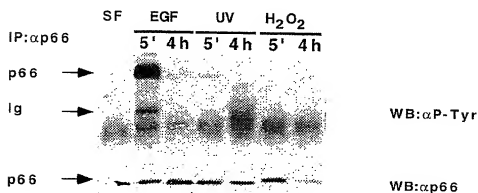
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Figure 1

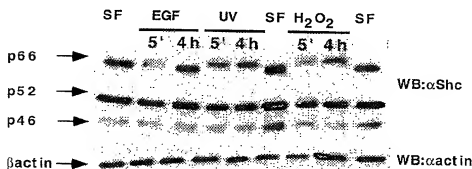
A



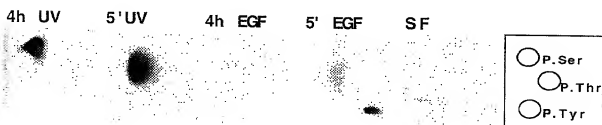
B



C

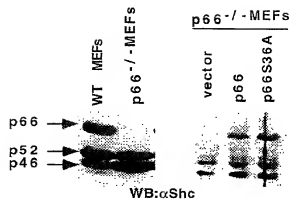


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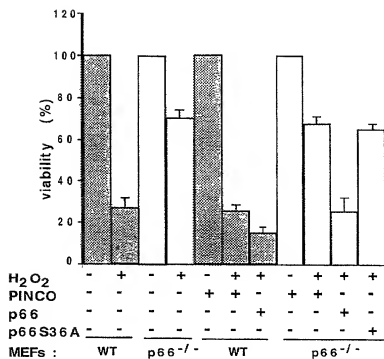


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Figure 2

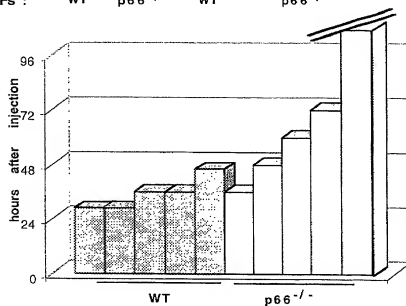
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B

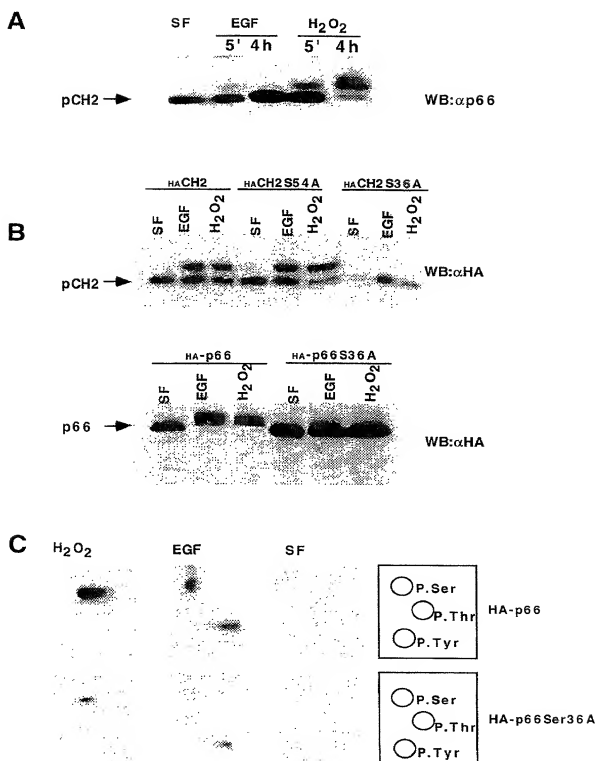


C



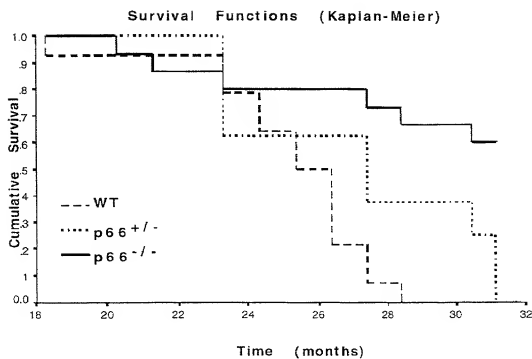
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Figure 3



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Figure 4



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Figure 5a

10	20	30	40	50	60
atggggcctg	aaactgtctg	ggctctgagct	ggggagcggg	agccacttgt	ccctctccct
70	80	90	100	110	120
ccccaggact	tctgtgactc	ctggggccaca	gaggtccaac	cagggttaagg	gcctgggggat
130	140	150	160	170	180
acccccctgc	tggccccctt	gccccaaactg	gcaggggggc	caggctgggc	agcagccccct
190	200	210	220	230	240
ctttcacctc	aactATGSA	CTCCTGCCCC	CCAAGCCCAA	GTACAATCCA	CTCGGAATG
250	260	270	280	290	300
AGTCTCTGTC	ATGCTGGAG	GAAGGGGCTT	CTGGGTCCAC	CCCCCGGAG	GAGCTGCTT
310	320	330	340	350	360
CCCCATCAGC	TTCATCCCTG	GGGCCCATCC	TGCTCTCTCT	GCCTGGGGAC	GATAGTCCCA
370	380	390	400	410	420
CTACCTCTGT	CTCCTCTCTC	CCCCGGATGA	GCAACTTGAG	GCTGGCCAAC	CCGGCTGGGG
430	440	450	460	470	480
GGGCCCCAGG	GTCTAAGGGG	GAGCCAGGAA	GGGCAGCTGA	TGATGGGGAG	GGGATCGATG
490	500	510	520	530	540
GGGCAGCCAT	GCCAGASTCA	GGCCCCCTTAC	CCCTCTCTCCA	GGACATGAAC	AAGCTGAGTG
550	560	570	580	590	600
GAGGCGCGGG	GCCAGGACTT	CGGGTGAAG	GGGGCCAGCT	TGGGGGCGAG	GAGTGAACCC
610	620	630	640	650	660
GCCAGGGGAG	CTTTGTCAAT	AAGCCCAAGC	GGGGCTGGCT	GCATCCCAAC	GACAAAGTCA
670	680	690	700	710	720
TGGGACCCGG	GCTTTCCTAC	TTGGTTCCGT	ACATGGGTTG	TGTGGAGGTC	CTCCAGTCAA
730	740	750	760	770	780
TGCGTGCCTT	GGACTTCAAC	ACCCCGGACTC	AGGTCAACAG	GGAGGCCATC	AGTCTGGTGT
790	800	810	820	830	840
GTGAGGCTGT	GCCGGTGCT	AAGGGGGCGA	CAAGGAGGAG	AAAGCCCTGT	AGCGCGCCGC
850	860	870	880	890	900
TCAGCTCTAT	CTGGGGGAGG	AGTAACTTGA	AATTGTGCTG	AATGCCAATC	ACTCTCAACG
910	920	930	940	950	960
TCTCCACACG	CAGCCTCAAC	CTCATTGGCG	CAGACTTGCAA	ACAGATCATC	GCCAAACACC
970	980	990	1000	1010	1020
ACATGCACATC	TATCTCATTT	GCATCCGGCG	GGGATCCGGA	CACAGCCGAG	TATGTGACCT
1030	1040	1050	1060	1070	1080
ATGTTGCCAA	AGACCTCTGT	AATCAGAGAG	CCTGCCACAT	TCTGGAGTGT	CCCGAAGGGC
1090	1100	1110	1120	1130	1140
TTGCCACAGG	TGTCATCAGC	ACCATTTGGCC	AGGCCTTCGA	GTTCGCTTTC	AAACAATACC
1150	1160	1170	1180	1190	1200
TCAGGAACCC	ACCCAAACTG	GTCACCCCCTC	ATGACAGGAT	GGCTGGCTTT	GATGGCTCAG
1210	1220	1230	1240	1250	1260
CATGGGATGA	GGAGGAGGAA	GAGCCACCTG	ACCATCAGTA	CTATAATGAC	TTCCCGGGGA
1270	1280	1290	1300	1310	1320
AGGAACCCCC	CTTGGGGGGG	GTGGTAGACA	TGAGGCTTCG	GGAAGGAGCC	GCTCCAGGGG
1330	1340	1350	1360	1370	1380
CTGCTCGACC	CACTGCAACC	AATGCCCCAGA	CCCCAGCCGA	CTTGGGAGCT	ACATTGCTGT

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Figure 5a continued

1390	1400	1410	1420	1430	1440
TAGGACAGCC	TGTTGGGGGA	GATCCAGAAG	TCCGCAAAACA	GATGCCACCT	CCACCAACCT
1450	1460	1470	1480	1490	1500
GTCCAGGCAG	AGAGCTTTTIT	GATGATCCCT	CCTATGTCAA	CGTCCAGAAC	CTAGACAAGG
1510	1520	1530	1540	1550	1560
CCCGGCAAGC	AGTGGGTGGT	GCTGGGCCCC	CCAACTCTGC	TATCAATGGC	AGTGCAACCC
1570	1580	1590	1600	1610	1620
GGGACCTGTT	TGACATGAAG	CCCTTCGAAG	ATGCTCTTCG	GGTGCCCTCA	CCTCCCAAGT
1630	1640	1650	1660	1670	1680
CGGTGTCCAT	GGCTGAGCAG	CTCCGAGGGG	AGCCCTGGTT	CCATGGGGAAG	CTGAGCCGGC
1690	1700	1710	1720	1730	1740
GGGAGGCTGA	GGCACTGCTG	CAGCTCAATG	GGGACTTCTT	GGTACGGGAG	AGCAGCAACA
1750	1760	1770	1780	1790	1800
CACCTGCAACA	GTATGTGCTC	ACTGGCTTGC	AGAGTGGGCA	GCCTAAGCAT	TTGTACTATG
1810	1820	1830	1840	1850	1860
TGGACCTCTA	GGGTGTGGTT	CGGACTAAGG	ATCACCGCTT	TGAAAGTGTC	AGTCACCTTA
1870	1880	1890	1900	1910	1920
TCAGCTACCA	CATGGACAAT	CACCTTGCCA	TCACTCTCTC	GGGCAGCGAA	TGTGTCTTAC
1930	1940	1950	1960	1970	1980
AGCAACTGT	GGAGCGGAAA	CTGtgatctg	ccctagocgt	ctcttcacaga	agatgccttc
1990	2000	2010	2020	2030	2040
caatcctttc	cacctatttc	octaactctc	gggacctogt	ttggggagtgt	tctgtgggct
2050	2060	2070	2080	2090	2100
tggccttggt	tcagagctgg	gagtagcatg	gactctgggt	ttcatatcca	gctgagttag
2110	2120	2130	2140	2150	2160
aggggtttgag	tcaaaagcct	gggtgagaat	cctgcctctc	cccaaacatt	aatcaccaaa
2170	2180	2190	2200	2210	2220
gtattaatgt	acagagtggc	ccctcacctg	ggcctttcct	gtgccaacct	gatgcccttt
2230	2240	2250	2260	2270	2280
ccccaagaag	gtgagtgcct	gtcatggaaa	atgtcctgtg	gtgacaggcc	cagtgggaaca
2290	2300	2310	2320	2330	2340
gtcacccttc	tgggcaagg	ggaacaaatc	acacctctgg	gcttcagggt	atcccagacc
2350	2360	2370	2380	2390	2400
cctctcaaca	ccgcoccccc	ccatgtttta	actttgtgcc	tttgaccatc	tcttaggtct
2410	2420	2430	2440	2450	2460
aatgataatt	tatgcaaaaca	gttcttggac	ccctgaattc	ttcaatgaca	gggagtcaca
2470	2480	2490	2500	2510	2520
cacctctctg	gcttctggga	cctgtgttct	tgctgagcae	cctctccggt	ttgggttgagg
2530	2540	2550	2560	2570	2580
ataacagagg	caggagtggc	agctgtcccc	tctccctggg	gatatgcaac	ccttagagat
2590	2600	2610	2620	2630	2640
tgccccagag	ccccactccc	ggccaggcgg	gagatggacc	cctccctctg	tcagtgcctc
2650	2660	2670	2680	2690	2700
ctggccgggg	ccctccaccc	caaggggtct	gtatatacat	ttcataaggc	ctgcctctcc
2710	2720	2730	2740	2750	2760

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Figure 5a continued

atgtgtcatg	cctatgtact	ctgcgcctaaa	gtgcagccct	tctctctgaa	gcctctgcc
2770	2780	2790	2800	2810	2820
tgcttccctt	tctgggaggg	cggggtgggg	gtgactgaat	ttgggcctct	tgtagactta
2830	2840	2850	2860	2870	2880
actctcccat	gtggattttg	tggaggttag	aaaaggggca	ttgagactat	aaagcagtag
2890	2900	2910	2920	2930	2940
acaatcccca	cataccactc	gtagagttag	aactgcattc	ttttaaagtt	ttatatgcat
2950	2960	2970	2980	2990	3000
atatatttag	gctgtctagc	ttaatttctc	atattctctt	ccattgctta	ttcttgagca
3010	3020	3030	3040	3050	3060
caaaatgata	atcaattatt	acattttata	atcacctttt	tgacttttcc	aagccctttt
3070	3080	3090	3100	3110	3120
acagctcttg	gcatttttct	cgcctaggcc	ttgtaggtaa	ctgggactgc	accttttata
3130	3140	3150	3160	3170	3180
ccagagacct	gaggcagatg	aaatttattt	ccatctagga	ctagaaaaac	ttgggtctct
3190	3200	3210	3220	3230	3240
taccgcgaga	ctgagagcca	gaagtacgcc	cgaatgcctg	tcagtttcat	ggaggggaaa
3250	3260	3270	3280	3290	3300
cgcaaaacct	gcagtttctg	agtacctctc	acaggcccg	cccagcctag	gccccgggtg
3310	3320	3330	3340	3350	3360
gccacaccac	agcaagccgg	ccccctctc	tttgctctc	tggaataagg	agagttagcc
3370	3380	3390	3400	3410	3420
gttttctatc	tggtctctct	ttgctgtttg	gatgtttcca	cgggtctcac	ttataccaaa
3430	3440	3450	3460	3470	3480
gggaaaaact	tteattaaag	tcctgtattc	ttctaaaaaa	aaaaaaaaaa	aaatacattt
3490	3500	3510	3520	3530	3540
atacatacc	tttttgactt	ttccaagccc	ttttacagct	cttggcattt	tctctgccta
3550	3560	3570	3580	3590	3600
ggcctgtgag	gtaactggga	tcgcactttt	tataccagag	acctgaggca	gatgaaattt
3610	3620	3630	3640	3650	3660
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3670	3680	3690	3700	3710	3720
agcc.....

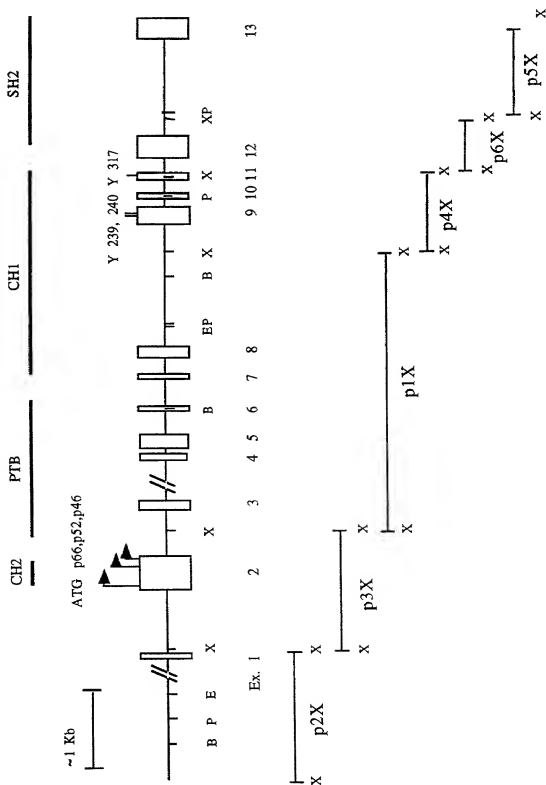
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Figure 5b

10	20	30	40	50	60
MDLLPPKPKY	NPLRNESLSS	LEEGASGSTP	PEELPSPSAS	SLGPILPPLP	GDDSPITLCS
70	80	90	100	110	120
FFPRMSNLRL	ANPAGGRPGS	KGEPRGAADD	GEGIDGAAMP	ESGPLPLLD	MNKLSGGGGR
130	140	150	160	170	180
RTRVEGGQLG	GEEWTRHGSF	VNKPTRGWLH	PNDKVMGPGV	SYLVRVMGCV	EVLQSMRALD
190	200	210	220	230	240
FNTRTQVTRE	AISLVCEAVP	GAKGATRRRK	PCSRPLSSIL	GRSNLKFAGM	PITLTVSTSS
250	260	270	280	290	300
LNLMADCKQ	IIANHHMQSI	SFASGGDPDT	AEYVAYVAKD	PVNQRACHIL	ECPEGLAQDV
310	320	330	340	350	360
ISTIGQAFEL	RFKQYLRNPP	KLVTPHDRMA	GFDGSAWDEE	EEEEPPDHQYY	NDFPGKEPPL
370	380	390	400	410	420
GGVVMRLRE	GAAPGAARPT	APNAQTPSHL	GATLFGQPV	GGDPEVRKQM	PPPPPCPGRE
430	440	450	460	470	480
LFDDPSYVNV	QNLDKARQAV	GGAGPPNPAL	NGSAPRDLFD	MKPFEDALRV	PPPPQSVSMA
490	500	510	520	530	540
BQLRGEPIWFH	GKLSRREAEA	LLQLNGDFLV	RESTITPGQY	VLTGLQSGQP	KHLLLVDPGE
550	560	570	580		
VVRTKDHRFE	SVSHLISYHM	DNHLPIISAG	SELCLQQPVE	RKL*	

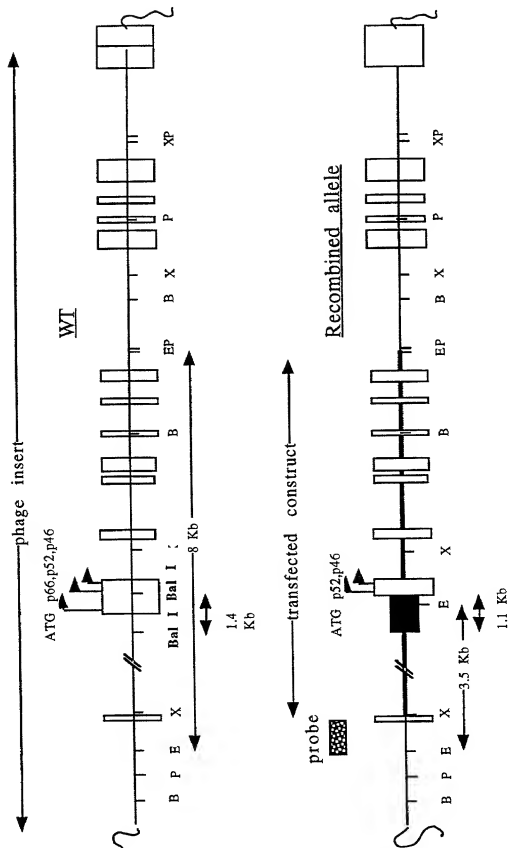
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Figure 6



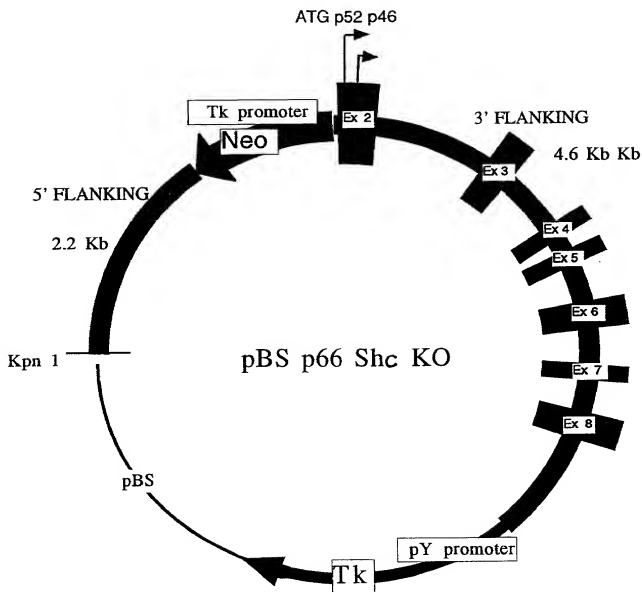
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Figure 7

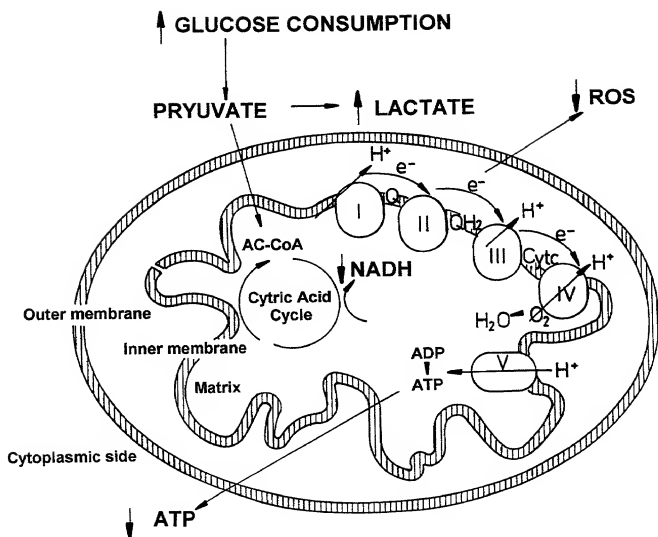


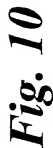
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Figure 8

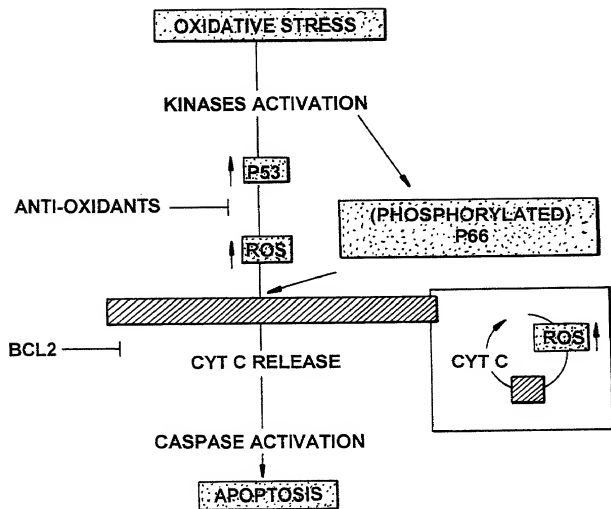


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*Fig. 9*



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*Fig. 11*